

Software Manual

ZEISS Labscope v3.4



ZEISS Labscope v3.4

Original Manual

Carl Zeiss Microscopy GmbH Carl-Zeiss-Promenade 10 07745 Jena Germany microscopy@zeiss.com www.zeiss.com/microscopy

Carl Zeiss Suzhou Co., Ltd. Modern Industrial Square 3-B, No.333 XingPu Road SIP 215126 Suzhou China

Document Name: Software Manual ZEISS Labscope v3.4 Revision: 1 Language: en-US Effective from: 06/2022

© 2022 Without the prior written consent of ZEISS, this document or any part of it must neither be translated nor reproduced or transmitted in any form or by any means - including electronic or mechanic methods, by photocopying, recording or by any information or filing system. The right to make backup-copies for archiving purposes shall remain unaffected thereby. Any violations may be prosecuted as copyright infringements.

The use of general descriptive names, registered names, trademarks, etc. in this document does not imply that such names are exempt from the relevant intellectual property laws and regulations and therefore free for general use. This shall also apply if this is not specifically referred to. Software programs shall entirely remain the property of ZEISS. No program or subsequent upgrade thereof may be disclosed to any third party, copied or reproduced in any other form without the prior written consent of ZEISS, even if these copies or reproductions are destined for internal use at the customer's only, the only exception being one single back-up copy for archiving purposes.

Content

| 1 | Introduction to Labscope | 5 |
|---------------------------------------|---|--|
| 2 | Installing Labscope | 6 |
| 3 | First Start & Overview | 7 |
| 4 | Microscope Configuration | 8 |
| 5 | Multitask Bar | 10 |
| 6 | Live View | 11 |
| 7 | Advanced Tools in Live View | 12 |
| | 7.1 Graticule Overlay | 13 |
| | 7.2 Over Exposure Indicator | 13 |
| | 7.3 Focus Indicator | 14 |
| | 7.4 Split View | 14 |
| 8 | Image View | 16 |
| | | |
| 9 | Advanced Tools in Image View | 18 |
| 9 1(| Advanced Tools in Image View 0 Histogram View | 18 19 |
| 9 1(1 ⁻ | Advanced Tools in Image View 0 Histogram View 1 Image & Video Acquisition | 18 19 20 |
| 9 1(1 ⁻ | Advanced Tools in Image View 0 Histogram View 1 Image & Video Acquisition | 18 19 20 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition 11.1 Acquisition Modes. 11.1.1 Fast Panorama Mode. | 18 19 20 20 22 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition 11.1 Acquisition Modes. 11.1.1 Fast Panorama Mode. 11.2 Acquiring Multi-Channel Images. | 18 19 20 20 22 23 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition | 18 19 20 20 22 23 25 |
| 9 1(1 ⁻ | Advanced Tools in Image View 0 Histogram View 1 Image & Video Acquisition 11.1 Acquisition Modes 11.1.1 Fast Panorama Mode 11.2 Acquiring Multi-Channel Images 11.3 Acquiring EDF Images 11.4 Recording Time Series Videos | 18 19 20 20 21 22 23 25 26 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition | 18 19 20 20 21 22 23 25 26 27 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition . 11.1 Acquisition Modes. 11.1.1 Fast Panorama Mode. 11.2 Acquiring Multi-Channel Images. 11.3 Acquiring EDF Images. 11.4 Recording Time Series Videos . 11.5 Acquiring Fast Panorama Images. 11.6 Acquiring images with confluency and counting result | 18 19 20 20 20 21 22 23 25 26 27 28 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition 11.1 Acquisition Modes. 11.1.1 Fast Panorama Mode. 11.2 Acquiring Multi-Channel Images. 11.3 Acquiring EDF Images. 11.4 Recording Time Series Videos 11.5 Acquiring Fast Panorama Images. 11.6 Acquiring images with confluency and counting result 11.7 Acquisition Settings | 18 19 20 21 22 23 26 26 26 26 26 26 26 27 28 30 |
| 9 1(1 ⁻ | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition . 11.1 Acquisition Modes. 11.1.1 Fast Panorama Mode. 11.2 Acquiring Multi-Channel Images. 11.3 Acquiring EDF Images. 11.4 Recording Time Series Videos . 11.5 Acquiring Fast Panorama Images. 11.6 Acquiring images with confluency and counting result . 11.7 Acquisition Settings. 11.8 File Name Template Configuration . | 18 19 20 20 20 21 22 23 25 26 27 28 30 31 |
| 9 1(1 [,] | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition | 18 19 20 21 22 23 26 26 26 26 26 27 30 31 32 |
| 9 1(1 [°] 1: 1: | Advanced Tools in Image View. 0 Histogram View. 1 Image & Video Acquisition | 18 19 20 21 22 23 26 26 26 26 26 27 30 31 32 32 34 |

| 13.2 Generating Reports for Multi-Channel Images | . 36 |
|---|------|
| 14 Files View | 38 |
| 15 Image Processing | 39 |
| 16 Global Settings | 40 |
| 16.1 Licensing via Free Trial, purchase or activation | . 41 |
| 17 Application and System Requirements | 43 |

1 Introduction to Labscope

Labscope is your easy-to-use imaging app for connected microscopes. Be it in the laboratory, university, school or even for your hobby – with Labscope you snap images, record videos and measure your microscopic samples, easier than ever. Transform your ZEISS network-compatible microscopes into Wi-Fi-enabled imaging systems. You can easily create digital classrooms or digital labs. Connect to microscopes at any point in time, from anywhere within your room. And then, share your images with the touch of your fingers.

Digital Classroom With the imaging app Labscope you can transform your student lab into a digital classroom via a WiFi-enabled network. Simply integrate your microscopes into the network - and you're ready to display all the live images at any time and from anywhere in the room in real-time. Project images onto the wall or a TV screen and foster teamwork among the students. With Labscope 'lecture & listen' becomes 'show & see'.

- Routine
ApplicationsLabscope transforms your ZEISS network-compatible microscopes into a WiFi-enabled imaging
system. You can easily create digital labs just connect to any of the microscopes at any point in
time and from anywhere in your room. Be it in your histology, cytology, hematology or pathology
lab Labscope snaps images and records videos of your microscopic samples. You even acquire
multi-channel fluorescence images easier than ever.
- Windows, iOS & Whether you're using a Windows PC with mouse and keyboard, a Windows tablet with touch-Android Support Screen, an iPad, iPhone or Android devices – Labscope runs on all of these devices. Enjoy live images and view live thumbnails from your connected microscopes, and easily switch between all microscopes in the room. And snap images with scale bar and meta data. You can additionally perform annotation and measurement tasks. Record time-lapse or regular videos and save your images and videos to any shared network folder.
 - Service &
ContactYou can use the ZEISS Labscope Forum
cludes the FAQs and latest release information. If you have any ideas for Labscope development,
share with us in the ZEISS Portal My Voice.
 - Legal Notes Not all products are available in every country. Use of products for medical diagnostic, therapeutic or treatment purposes may be limited by local regulations.

Info

This manual is primarily based on Windows Labscope v3.4. And the latest iOS Labscope version is v3.3 and Android Labscope v4.0. Note there are certain functional differences between iOS, Android and Windows version of Labscope. For example, the Android and iPhone Labscope has less functions than iPad and Windows Labscope. You will find a detailed overview of the different functions in the product info under https://www.zeiss.com/labscope.

2 Installing Labscope

For Windows

- 1. Download the latest **Labscope v3.4** for Windows via the product website: <u>https://www.zeiss.com/labscope</u>
 - \rightarrow You will be directed to the <u>ZEISS portal</u> for downloading the installation files.
- 2. Double-click on LabscopeSetup_vx.exe to install the software.
- 3. Perform the required steps shown by the installation wizard. Please agree if you are asked to install additional drivers.
 - → Some modules of Labscope require additional installation, e.g. **BioModuleSetup.exe**, which is for **AI Cell Counting and AI Cell Confluency** models, and you can find the installer in the same page of Labscope download in Zeiss Portal.

After the installation you see the Labscope program icon on your desktop.

For iOS

Download and install the latest Labscope v3.3 for iOS via the App store. And you could choose the corresponding app for iPhone and iPad ffrom there.

Please visit our product website to find the links to the downloads in the App Store:

https://www.zeiss.com/labscope

For Android

Download and install the latest Labscope v4.0 for Android via Google website to find the links to the downloads:

https://www.zeiss.com/labscope

3 First Start & Overview



Click on the app icon on your device to start the app.

Establish connection for the Labscope device with your microscope(s) and camera(s).

- For iPhone and Android devices, WiFi connection is supported.
- For iPad, WiFi or ethernet connection via lightning or USB-type-C adapters is supported.
- For Windows, WiFi, USB and ethernet connection is supported.

Note: Different microscopes and cameras support different connectivity as well.

After connection is established, the microscopes will be automatically shown up in Labscope. When there are no microscopes found in current connection, Labscope will show virtual microscopes. The below image is an example after connecting with several microscopes at same time.



4 Microscope Configuration



Fig. 2: Microscope Configuration

1 Select Microscope/Camera

Select the connected microscope or camera from the list. If the microscope is encoded, the microscope will be recognized automatically.

2 Enter Parameters and Perform a Shading Correction

This fields allow you to enter a name for the microscope and select the camera adapter.

Click Setup to open the Shading Correction Setup.

We recommend to perform a shading correction for each objective of any newly configured microscope before you start to work with the device.

Note: When working in the shading correction setup window, you need to remove the sample out of the live view for transmitted light or use a mirror for reflected light. Please read the on-screen hints in Labscope carefully. The shading correction is correlated to each objective, which means wrong selection of an objective in the setup window or live view can lead to a wrong live image effect.

3 Lock Configuration

If activated, you can enter a 4 digits PIN to lock the microscope configuration. If you forget the PIN code, the universal PIN code to unlock is: h!%jPYtt34

4 Select Objective/Filter set

Depending on the connected microscope the available positions (e.g. 1-6) will be displayed. If you click on a position you can select the attached component (e.g. 5 x objective) from the list.

5 Scaling calibration

In manual calibration mode drag the measurement line to a known size object from your sample. To finish the scaling calibration of the objective, input the correct length. If you have a ZEISS standard calibrator, you can also use Auto Calibration mode to easily calibrate.

Note: The calibration is correlated to each objective. There is an indicator on a calibrated objective in the Microscope Configuration window. A wrong selection or changing the camera adapter selection can lead to wrong scaling info.

6 Only use this microscope

If activated, only the current microscope is displayed in the microscope view.

5 Multitask Bar

The Multitask bar is the central element for navigating through the different views of the app. It can be accessed from the top toolbar on any screen of the app. It allows to easily switch between the different views by simply clicking on the desired thumbnail / preview window. The following views are available and will be explained in the next chapters:

- Microscope View, see First Start & Overview [> 7].
- Live View, see Live View [> 11].
- Image View, see Image View [> 16].
- File View, see Files View [> 38].



Fig. 3: Multitask bar

1 Thumbnail View of the available views

Click on the corresponding thumbnail to change to the desired view.

6 Live View

The Live view shows the live image from the connected camera on the selected microscope.

Fig. 4: Live View

1 Snap Button

Acquires an image (or short a "Snap"). Depending on the selected acquisition mode you can perform different types of acquisition.

2

Change Acquisition Mode

Select the desired acquisition mode, see Acquisition Modes [> 20].

3 Display of Objective & Reflector

Shows the current objective and reflector. When working with non-encoded microscopes you can select the corresponding components manually. For encoded microscopes, the currently used objective and reflector will be displayed automatically.

4 Camera Acquisition Settings

Opens the camera acquisition settings, see *Acquisition Settings* [> 30].

5 Histogram

Opens the histogram panel, see *Histogram View* [> 19].

6 Show More Tools / Advanced Tools

Displays more tools (also called "advanced tools") in the toolbar, see *Advanced Tools in Live View* [> 12].

For all available annotations and measurement tools, see *Annotations and Measurement Tools* [> 32].

7 Advanced Tools in Live View

In **Live** view, click the **Show More Tools** button at the bottom right corner to see more tools. The additional tools / functions are displayed in the main toolbar



The following functions are available:

| lcon | Function | Description | |
|-----------|---|---|--|
| ŧ | Annota- tions and Measure- ment Tools | This function is available for both Live view and image view, see <i>An-notations and Measurement Tools</i> [> 32]. | |
| \oplus | Graticule Overlay | Add various overlays to the image, see <i>Graticule Overlay</i> [> 13]. | |
| | Over Ex- posure In- dicator | Displays areas with over exposure, see <i>Over Exposure Indicator</i> [> 13]. | |
| <u>()</u> | Focus In- dicator | Displays a focus indicator bar, see <i>Focus Indicator</i> [> 14]. | |
| ++ | Split View | Opens the Split (Comparison) view, see <i>Split View</i> [> 14]. | |
| * | Laser Pointer | This function is available for both Live view and image view. It will be shown when the Education module is activated. | |
| | | Displays a laser pointer/ red dot. It will follow the mouse or finger movement on the screen. This is useful for teaching or presentation purposes. | |

| lcon | Function | Description |
|------|-----------------|---|
| 1 | Drawing Mode | This function is available for both Live view and image view. It will be shown when the Education module is activated. |
| | | Displays a combined view of both microscopy view and camera view. This can be useful for drawing sketches from a sample image. |
| | | We recommend to use a tablet holder (not included) for the drawing process. |

7.1 Graticule Overlay

Graticule Overlay

You will see 4 windows where you can select the following overlays: no graticule, grid, cross with divisions and crosshair. Select the desired overlay by clicking on it.



7.2 Over Exposure Indicator

This function is available for both live view and image view. After clicking, the regions having over exposure will be shown with red color.



7.3 Focus Indicator

After clicking on the icon in the main toolbar, the focus indicator bar will show up at the central bottom of the **Live** view. When the focus changes, the indicator will also change dynamically.

You can even manually calibrate the default status of the focus indicator. Firstly adjust the fine focus of the microscope to make the image clear. Then click on the indicator bar and it will be calibrated.



7.4 Split View

This view is available for Live and Image view.

↔



After clicking on the Split view icon, two panels are displayed initially. The left side shows the current view (Live or Image view) and the right side shows the files of Labscope.

By selecting another file from the list it will be displayed in a comparison view together with the image on the left side:



8 Image View

After you have acquired an image you can quickly switch to the image view by clicking on the thumbnail / preview image of the snap in the lower right corner of the image area. The thumbnail / preview image will be available for a few seconds so that you can continue with the image acquisition of another image without interrupt. You can open an image from the **File view** as well.



Fig. 5: Image View

Change Scaling

Shows the current image scaling. Click to change the scaling by selecting another image.

Annotations and Measurement Tools 2 Shows the available annotations and measurement tools which can be added to the image, see chapter Annotations and Measurement Tools [> 32]. 3 Histogram Shows the histogram panel. Edit File Name 4 Either change the file name or configure the file name template. 5 Previous/ Next Image Click < to go to the previous image. Click > to go to the next image. 6 Share Image Shares the image directly via email or other tools. Save Image 7 Saves the image with different formats.

Shows additional tools, e.g. image processing, over exposure indicator and Split view. See also chapter *Image Processing* [> 39].

9 Advanced Tools in Image View

In **Image** view, click the **Show More Tools** button at the bottom right corner to see more tools. The additional tools/functions are displayed in the main toolbar.



The following functions are available:

| lcon | Function | Description | |
|------|-----------------------------------|--|--|
| * | Image Process- ing | Displays the image processing panel. You can edit the value of gamma, brightness, contrast, color intensity and sharpness, see <i>Image Processing</i> [> 39]. | |
| Z | Over Ex- posure In- dicator | Displays areas with over exposure, see <i>Over Exposure Indicator</i> [> 13]. | |
| ++ | Split View | Opens the Split (Comparison) view, see <i>Split View</i> [> 14]. | |
| Ē | Multi- channel | This function will be available and active only when the Multi-Chan- nel module is activated and a multi-channel image was opened. | |
| | son | Shows all individual channel images in separate windows. The last window displays the combined multi-channel image. | |

See also

Annotations and Measurement Tools [32]

10 Histogram View

This function is available for both **Live** view and **Image** view. Click the button in main toolbar to open the histogram panel.



1 Color Range

Use the **Min/Max** and **Best Fit** buttons to achieve a proper display effect just by one click

2 Gamma

Adjust the gamma value by clicking, dragging or inputting.

3 Advanced Options

Adjust the histogram value of black and white by dragging the scroll bar or inputting in the dialog boxes

Info

For JPEG or TIFF format, the black and white value will be 0 and 255 (example for 8 Bit depth images) after the histogram is saved. Note that the "**Min/Max**" indicator in histogram panel for JPEG and TIFF is highlighted in most cases.

11 Image & Video Acquisition

Image acquisition is directly done in the **Live View**. First you have to select the desired acquisition mode. Per default, the single image acquisition mode is selected.

Note: Availability of each acquisition mode is depending on camera and microscope types and also licensed modules.



1 Acquisition Mode Selection

Select the desired acquisition mode, see Acquisition Modes [> 20].

2 Acquisition/ Camera Settings

Opens the acquisition settings. See also chapter Acquisition Settings [> 30].

See also

Acquiring Multi-Channel Images [> 23]

11.1 Acquisition Modes

The following acquisition modes are available and can be selected by tapping on the corresponding mode icon on the right side:

| lcon | Mode | Description |
|------|------------------------------------|--|
| 0 | Single Image Acquisition (Snap) | Acquires a single image (also called a Single Snap). |
| ā | Fast Snap | Acquires an image from live view at high capture speed. In this mode, multiple users can acquire an image simultane-ously. |
| | Multi Channel Acquisition | Only available if you have licensed the Multi Channel mod- ule. |

| lcon | Mode | Description |
|--------|---|---|
| | | Acquires fluorescence and transmitted light images in independent channels. The module supports adding false-color, comparing channels and creating reports with display of individual channels, see <i>Acquiring Multi-Channel Images</i> [> 23]. |
| EDF | Extended Depth of Field (EDF) Acquisition | Acquires an EDF image, see <i>Acquiring EDF Images</i> [> 25]. |
| ഘ | Video Recording | Records a video. This functionality allows to record the cur- rent live image as a movie. |
| С С | Time Series Recording | Records a time series video, see <i>Recording Time Series Videos [</i> 26]. |
| | | This functionality allows to set up time intervals (e.g. each 5 seconds) where an image is taken. The images are then combined to a video. |
| ¢ | Fast Panorama | Only available if you have licensed the Fast Panorama module for Windows Labscope since v3.3. Acquire whole slide images with your manual microscope, see <i>Acquiring</i> <i>Fast Panorama Images</i> [> 27]. |
| E. | Bio Modules | Only available if you have licensed the AI Cell Confluency and/or AI Cell Counting modules for Windows Labscope since v3.4. Acquire images and get the confluency and/or counting result based on the live view, see <i>Acquiring im- ages with confluency and counting result</i> [> 28]. |

Note that there can be more types of snapping modes which are only available for certain types of cameras e.g.:

is for a denoised snap.

See also

Acquisition Settings [> 30]

11.1.1 Fast Panorama Mode

1 Hints

Hints show up during Fast Panorama acquisition. They will show the information of how many images are stitched, the current whole image resolution and current FPS data.

2 Preview of the stitched panorama image

The red box shows the current position of live view.

3 Live view window with focus indicator

The outline color of the live view has two colors with different meanings. White is the normal status with successful stitching. Yellow means the confidence level of the current stitching attempt is low or the current image is unable to be stitched. This can be caused by no matching area, or the matching area is too small. Once it completely lost position or cannot stitch, a reminder occurs above the live view for reminding users to move it back slowly. The bar below the live window is the focus indicator which shows the calculated focus status. During the stage moving and stitching process, a blue outline always shows in the background which indicates the last successfully stitched position.

4 Stitched images as background

Position search This button is enabled when the position is completely lost. After clicking on the button, the live view image will be searched from the whole stitched image background. If the correct position is found, Labscope will route to the correct position. If not, please try to move the live view to other areas where may be an overlap with the background, then try gain.

Undo Click to delete the last stitched image. It supports continuous clicks, then multiple stitched images will be deleted.



Cancel Click to stop the panorama acquisition.

~

Save Click to save the panorama result.

5

Note: For more information about the acquisition see Acquiring Fast Panorama Images [> 27].

11.2 Acquiring Multi-Channel Images

Labscope provides an easy solution for multi-channel acquisitions. Enhanced functions are available, like editing pseudo color, one-key multi-channel acquisition, comparison view, and report with displaying each channels, etc..

Note: Currently the multi channel module function is available for Labscope for iPad and Windows. The supported camera types are: Axiocam 202 mono, Axiocam 208 color, Axiocam 105 color (Windows only), Axiocam 305 mono/color (Windows only).



1. Select Multi-Channel acquisition mode.

2. Click on the corresponding channel button to select the channel you want to snap.



- → If it is an encoded Axioscope 5 with Colibri light source, the light is turned on when the channel is activated.
- → If it is an encoded Axiolab 5, the activated channel is synchronized with the light source.
- → If it is a non-encoded microscope, the channel can only be activated manually on the microscope.

Note: The availability of channel buttons is dynamically changing according to the filter set. E.g. for a single DAPI bandpass filter, only the 385 LED channel will be displayed. The mapping logic is according to the excitation range of filter set and the LED's wavelength.

3. Activate the pseudo color panel of the current channel by long pressing on the individual channel button (for iPad). Or:

Right click on the individual channel button (for Windows). Then select the pseudo color by clicking on it.

Note: By clicking the button **III** in the main toolbar you can switch the image view between false color and b/w mode.



- 4. Adjust the acquisition, histogram settings and channel name for the current channel. The acquisition settings and channel name will be stored in Labscope.
- 5. Click **Snap** to acquire the image for selected channel.
 - → The preview of multi-channel image will be shown on the bottom right. It also contains 3 buttons (**Delete/Preview/Finish**).



- 6. Acquire the images for other channels.
- 7. After the acquisition for all needed channels is done, click **Finish**.
 - → The multi-channel image will be saved.

One-key/Manual Mode

Once a multi-channel acquisition is finished in Labscope by using encoded Axiolab 5 or encoded Axioscope 5, the acquired channels and each channel's acquisition settings will be stored as a multi channel template in Labscope. Additionally the **One-key/Manual** buttons will show up in Live view/Multi-Channel mode.



- In Manual mode you can acquire multi-channel images step by step manually.
 Note: You can always go back to any channel and overwrite it with a new snap before you click on Finish. After each manual acquisition, the multi channel template will be overwritten by the new template.
- In **One-key** mode an automatic acquisition will be performed, which is just following the last stored steps and settings. The handling process of this function differs between different microscope types and the connection types.

Stored data for each enabled channel:

- Exposure mode and exposure time
- Gain value
- Light intensity

Note: To use the One-Key functionality for **Axiolab 5 FL**, you have to switch the channel manually on the microscope. A message will appear: "Please switch to channel...".

11.3 Acquiring EDF Images

For samples with a different depth of field, it can be difficult to acquire a clear image for the whole area. In that case we recommend the EDF (Extended Depth of Field) acquisition mode to acquire combined in-focus images with different depths of field.

Note: The EDF acquisition only works with compound microscopes. It is not suitable for use with stereo microscopes.

- 1. Select **EDF** acquisition mode.

- Adjust the focus for a certain area and click Snap.
 → A image is acquired and a preview window pops up.
- 3. Adjust the focus and acquire another image. You can repeat this procedure until all areas are acquired in focus.
- 4. Click Finish to combine the acquired images to an EDF image.



11.4 Recording Time Series Videos

For viewing a slow changing dynamic sample, e.g. to observe slow movement, gradual evolution or morphology changes in a certain period of time, the **Time Series** (or Time Lapse) is a good acquisition mode to use.



1. Select Time Series acquisition mode.

- 2. Click Snap.
 - \rightarrow The Time Series settings dialog will be displayed.
- 3. Select the preferred recording settings (see Time Series Recording Settings) and click **OK**.
- 4. Click **Snap** again to finish the recording.
 - ightarrow The output time series video is generated and stored in the file system.

11.5 Acquiring Fast Panorama Images

The module **Fast Panorama** allows for easy acquisition of whole slide images (WSI) with manual microscopes. By manually moving the stage, images of the specimen will be stitched together automatically into a panoramic image. Please see the following steps for acquiring Fast Panorama images.

Note: Currently the module is only available for Windows since Labscope v3.3. The supported camera types are Axiocam 305 mono/color.

- After putting your sample on the microscope, adjust the focus to see a clear view. Set the correct objective in microscope configuration and set the shading correction accordingly, see *shading correction* [▶ 8].
- 2. In acquisition settings, choose manual exposure (the recommendation is below 2ms) and manual white balance. You may need to increase the light intensity and gain to a certain level to get a good brightness.
- 3. Select Fast Panorama mode 44. Move the sample to the desired position, then click the button again to start Fast Panorama acquisition.
- 4. Move the stage, the live images will be stitched together to a panorama picture. There are some hints and tools to support your panorama acquisition process, see below.
- 5. After finishing the Fast Panorama acquisition, click to finish and save your panorama picture. The image will be saved to your folder. You can adjust the histogram, do annotations and measurements, or save as JPG or TIF in the image view.



Important tips for getting a good panorama result:

- A good performance Laptop/PC is the basis for the performance of Fast Panorama process, see the required PC spec for Fast Panorama [*Application and System Requirements* [> 43]]
- Apply proper shading correction as preparation.
- Use manual exposure and manual white balance.
- Use low exposure time. Exposure time below 2ms is recommended, the lower the better performance.
- When moving the live view, having a certain overlap with the surrounding stitched images will improve the stitching quality.
- Try to use the tools Position search and Undo to support your process.
- For dense structure samples, e.g. tissue and blood, you could use a higher magnification objective for stitching once you meet the situation of frequent popup of the yellow box. Go from 10x to 20x, or from 20x to 40x, etc.

11.6 Acquiring images with confluency and counting result

The two Labscope modules **AI Cell Confluency and AI Cell Counting** are AI based solutions that automatically determine the confluency level of the adherent cell culture, and the number of adherent cells in the field of view, respectively. They deliver repeatable, objective results and do not require manual parameter adjustments.

Info

You need to install Labscope, then install **BioModules.exe**. Both are available to download from ZEISS Portal. Currently this function is only available for Windows Labscope with v3.4 or higher. The modules are primarily optimized for phase contrast images of adherent cell cultures.

1. In the live view, click **Bio Function**



2. Select the corresponding desired running models from the dialog.



- 3. Snap the desired view of your sample.
 - \rightarrow The confluency and/or counting result with the image will be simultaneously displayed.
- 4. Move the field of view to other areas of the sample and acquire multiple images if necessary.
 - \rightarrow There is always a preview window in the corner that immediately displays the results.
- 5. For enlarged preview, click **Zoom-In**



- → In the larger preview, the colors and transparency of confluency and counting masks can be edited.
- \rightarrow You can delete a particular selected scene and continue the workflow by going back to snapping.



6. To save your data, click Checkmark

in the small preview.

- → In Image View, you can check the confluency and/or counting masks and the data. You can also export the data table in csv format from the image.
- → The bio function also supports to trigger AI confluency/counting from an already acquired image.

11.7 Acquisition Settings

Based on the camera type and microscope system, the content of acquisition settings may vary. Using the **Reset all** button you can restore the default settings. When you enter the acquisition settings, the basic settings are displayed. When you click on **Advanced**, the advanced settings are displayed.

Basic Settings

| Parameter | | Description |
|-----------|---------------|--|
| Exposure | | |
| _ | Auto / Manual | Switch between manual and automatic mode for exposure time. When switched to manual mode, you can adjust the exposure time settings like Time and Gain manually. |
| _ | Intensity | Adjust the auto exposure target intensity. |
| _ | Push | Use the current target intensity and applies auto exposure once. |
| Wh | ite Balance | |
| - | Push | Apply auto white balance adjustment once. |
| _ | Auto / Manual | Switch between manual and automatic mode for white balance. When switched to manual mode, you can adjust the white balance settings manually using the Cold / Warm slider. |
| Lig | ht Intensity | Only available for AxioLab 5 and Axioscope 5 + Axiocam 202 mono / 208 color. |
| _ | Slider | Adjust the intensity of the light source. |
| _ | RL / TL | Switch the light path between reflected light (RL) and transmitted light (TL). |

Advanced Settings

| Parameter | Description |
|---|---|
| Microscope Name | Shows the microscope name. |
| MAC Adress | Shows the MAC address of the connected microscope. |
| IP Adress Shows the IP address of the connected microscope. | |
| Firmware Version | Shows the version of the currently installed firmware of the camera. |
| Live Quality | Adjust the settings for the live image quality here. |
| Image Orientation | Adjust the image orientation. |
| Color | Switch between Color mode and Grey mode for the color camera. |
| Gamma | Adjust the gamma settings. |
| Denoise | If enabled, denoising is automatically applied on the image. |
| Sharpening | If enabled, sharpening is automatically applied on the image. |
| Pixel Correction | If enabled, pixel correction is automatically applied on the image. |

| Parameter | Description |
|--------------------------------|--|
| HDR (High Dynamic Range) | If enabled, HDR is automatically applied on the image. |

Note: The availability of these settings is according to the camera types.

11.8 File Name Template Configuration

Labscope provides the possibility to configure individual file names before and after acquisition. If you click on the **Pencil** button in the top left corner of an opened image you can access the Configuration menu for file names. Several Parameter will be available:

| Parameter | Description | |
|---------------------------------|---|--|
| Configure File Name Template | Open the file name template configuration dialog. There you can se- lect or add templates on the left side. The following 5 elements for configuring a file name template are available: Text , Auto Numbe r, Date Time , Microscope Name and Objective Magnification . | |
| | If you activate Ask on save , every time when a snap is taken, a dia- log will pop up and ask you whether you want to change the name for the snapped file. You can also scan a barcode of your sample to input the file name, either by using a barcode scanner (for Windows) or the iPad camera itself (for iOS). Therefore you need to activate the parameter Camera-based barcode reader in the Global Settings menu, see <i>Global Settings</i> [> 40]. | |
| Auto Rename | Reset the file name according to the current file name template. | |

12 Annotations and Measurement Tools

You can add measurements, markers or text annotations either to the live image or an acquired image. The following list shows the available annotations and measurement tools:

| Parameter | Description |
|--------------------------|---|
| Distance | Draws a line and measures the length. |
| Rectangle | Draws a rectangle and measures area and perimeter. |
| Circle | Draws a circle and measures the area, perimeter and diameter. |
| Polygon | Draws a polygon and measures the area and perimeter. |
| Arrow | Draws an arrow. |
| Angle | Allows to measure an angle. |
| Count | Allows to draw in markers with ascending numbering. |
| Text | Allows to enter text in a box. |
| Scale bar | Allows to add a scale bar to the image. |
| Polyline | Draws a polyline and measures the distance. |
| Spline Contour | Draws a spline contour and measures the area and perimeter. |
| Spline | Draws a spline and measures the distance. |
| Disconnected Angle :ゴ | Draws a disconnected angle and measure the angle. |

| Parameter | Description |
|------------------|--|
| Caliper — | Draws a single caliper and measures the dis- tance. |
| Multi Calipers | Draws a multi-caliper and measures the dis- tances. |

See also

Global Settings [> 40]

13 Reports

13.1 Generating Reports

There are two ways to generate a report.

Generating a Report from Image View



→ A panel with **Report** and **Export as CSV** will be displayed.



- 2. Click **Report** to generate a PDF or RTF report based on selectable templates.
- 3. Click Export as CSV to export all measurements data from the image into a spreadsheet.

Generating a Report from Files View

- 1. In Files View, select multiple images at same time and click Report
 - Juliors
 Is all Constrained

 Is all Constrained
 Is all C
 - → No matter which way you have chosen to create the report, the window Select Report Template will show up.
 - Index

 Index
- 2. Select the desired template and click on it.

→ The window of **Input Missing Data** will be displayed.

圁





4. Click **Save** to store the report to the current files folder.

| isbure | - | | | | | | × |
|--|----|-------------------|---------|----|----|---|---|
| in the second second | | | × | ‡≡ | Ð, | G | |
| Shap 007.cst | | Report Preview | 5290 | | | | |
| MultipleChannels | -0 | | Autorit | | | | |
| brightficial Hodiney | | NOME: | | | | | |
| argentiela_vGanes | | | | | | | |
| Spot alumination | | | | | | | |
| HI Reflected light_b | | | | | | | |
| Rindlight_aphio_s | | | | | | | |
| spat Illumination state VE g/12/2020 | | | | | | | |
| apot Humination | | | | | | | |
| Verbiai Uuminat | | | | | | | |
| SE . | | s - spare (Vition | | | : | # | |
| i < i | | | | | | ٥ | |

13.2 Generating Reports for Multi-Channel Images

- 1. Activate Multi-Channel module.
- 2. Create a report as explained in chapter *Generating Reports* [> 34].



- 3. Select **Show channel comparison** to display the image of all channels (like in **Comparison / Split View**) in the report.
- 4. Select **Show individual channels** to display each individual channel image in the report.

6

14 Files View

In this view you can manage relevant files on your device.

| 1 | | 2 3 |
|--|-----------------|-------------------------|
| 📴 (uhrectau | | - <u>-</u> × |
| This PC > Ct > Dsers > 2CLZHAN9 > Documents > Test images | | t= 2 ⊡ ⊂4 |
| Image Video Report Tomplate | | Sort by name scienting |
| brightfield, Widney zar. Astococope 3. 02,001 | | Soft by name descending |
| Spot-Rumination_darkHeld_green-takewing_Stemi-305tit | | Sort by dato according |
| Rofloctod Nght Svightfield graan lecewing Stani 20519 | | Sort by date descending |
| Ringlight_aphild_StemN305_2.8H | | |
| spot illumination_ternspore_Stem1305dif | | |
| spot-Ulumination_vertical-Ulumination_dem-spores_Steml-305th realize subject units an | | |
| Vertical-Huminstor_pest_Sterni-305.07 | | |
| Vertifal Humisation past Stemi-335 1#F | | |
| Strap_005.czi | | |
| Snan COSkrei | | |
| | FreeSone (11)62 | |
| | | 0 |

Click to choose the locations. Only available for Windows version. Files for iOS and Android are stored in a default location.

2 Sort Files

Sort files according to the desired options.

- 3 Search Files
- 4 Open Folder

Opens the folder of the current file location.

- 5 Switch File Views
- 6 Delete / Share / Report / Info
- 7 Select All Files

15 Image Processing

Image processing can only be performed on an acquired single channel image. You can use the image processing functions to improve the display of the image according to your needs. The image processing functions are accessible via the **Show more tools** icon in the **Image View**. Then tap on the magic wand icon to display the available image processing functions. Note that currently image processing is not supported on multichannel images (*.czi format).



Fig. 6: Image Processing

1 Adjust Settings

Adjust the sliders of the individual functions to improve the image according to your needs.



Apply Settings or Cancel

16 Global Settings

| Parameter Description | | Description | | |
|-----------------------|--|--|--|--|
| General | | | | |
| – Language | | Select the language of the application. | | |
| | | Note : iOS and Android doesn't have this option here. The language will follow the system language settings. | | |
| _ | Show virtual mi- croscopes | If activated, the virtual microscope will displayed on the Microscope view screen. | | |
| _ | Microscopes can be added manually | If activated, you can manually add a microscope by typing in the IP address. Use this when microscopes are in different VLANs or subnets with the Labscope devices. | | |
| _ | Default file for- mat for snapped images | Choose the default image formats for image acquisition, e.g. CZI, JPG, TIF, CZI+JPG and CZI+TIF. | | |
| _ | Overwrite im- age on save | When saving an image, the previous image will be deleted. | | |
| _ | – Theme Select between dark and light theme. | | | |
| _ | Camera-based | Only available for iOS. | | |
| barcode reader | | Enables the scanning button to be shown in the file name window for Ask on save and searching text box. | | |
| _ | Hotkey for ac- quisition | Set the default acquisition hotkey for snapping and recording for the use of input devices like foot pedal or keyboard. If you use a foot pedal, you will need to set the hotkey of foot pedal in its driver to be same as the hotkey of Labscope. | | |
| | | tion hotkey is Space. | | |
| _ | Auto histogram acquisition | Set auto histogram (min/max or best fit) for each acquisition. The relevant histogram effect will be applied automatically for your acquisition each time. | | |
| Anr | notation | Different from the annotation adjustment in Live View or Image View, the settings here are for changing the default annotation properties. | | |
| _ | Measurement Unit | Change the unit of measurements here. | | |
| _ | Show measure- ments by de- fault | If activated, measurements are displayed by default when you add them to an image. | | |
| _ | Default color | Set the default color for annotations here. | | |
| _ | Transparent | If activated, the annotations are displayed with a transparent back- ground. | | |
| _ | Text back- ground color | Set the background color for text annotations here. | | |
| _ | Default size | Set a default text size here. | | |

| Par | ameter | Description | |
|-----|--|---|--|
| _ | Disable mea- surement color coding | Only available on iOS. A default random color is set when adding a new annotation. Activate this option, if you want to have the same color for each newly added annotation. | |
| - | Decimal places | Set the default decimal places for the measurements. | |
| Мо | dule Manager | Here you can enable/disable the corresponding modules. Note for some modules, there will be a hint showing that you have to restart the application to apply to the module changes. When you click Info , detailed module information will show up. Here you can also select Free trial or Purchase directly | |
| | Education | This module provides the functions of a Laser Pointer and Drawing | |
| _ | module (included) | Tube which are dedicated for the use of digital classrooms. In addi- tion, the module is also an access to the Labscope Teacher interface once the Labscope Teacher server is running. | |
| | | Note : Labscope Teacher is a licensed software. Consult your sales representatives for an enquiry, demo and free trial. | |
| _ | Multi Channel module | This module is enhanced with plentiful functions. It provides an easy solution for acquisition of fluorescence and transmitted light images in independent channels. It also supports adding false-color, comparing channels and reporting with displaying each channel. | |
| | | Note: The multi channel module function is available for Labscope for iPad and Windows. The supported camera types are: Axiocam 202 mono, Axiocam 208 color, Axiocam 105 color (Windows only) and Axiocam 305 mono/color (Windows only) | |
| _ | Fast Panorama module | This module allows for the easy acquisition of whole slide images (WSI) with manual microscopes. By manually moving the stage, im- ages of the specimen will be stitched together automatically into a panoramic image. | |
| | | Note: The module is only available for Windows Labscope since v3.3. The supported camera types are: Axiocam 305 mono/color. | |
| | Al Cell Conflu- ency module | Al based solution automatically determines the confluency level of the adherent cell culture. It delivers repeatable, objective results and does not require manual parameter adjustments. Note: The module is only available for Windows Labscope (v3.4 or higher). | |
| | AI Cell Count- ing module | Al based solution automatically determines the number of adherent cells in the field of view. It delivers repeatable, objective results and does not require manual parameter adjustments. Note: The module is only available for Windows Labscope (3.4 or higher). | |
| Pul | blisher | Displays legal information regarding the publisher as well as links to user support forum, data protection notice, and end user license agreement. | |

16.1 Licensing via Free Trial, purchase or activation

- 1. Under Global Settings, click Module Manager.
- 2. Click Free Trial or Purchase

→ Multiple choices will be displayed.

| meral | 6 | |
|------------|--|-------------------------|
| notations. | AI Cell Confluency | |
| blisher | Do you want to get the 30 days free trial or purchase the module? | Free Trial or Purchase |
| | | Free Trial or Funchase |
| | Get Trial Version | Freig Trial or Purchase |
| | Purchase | Free Trial or Purchase |
| | Activate | |
| | Cancel | |

- **Trial Version:** It will route to ZEISS Portal where you can apply for a 30-day trial license.
- **Purchase:** It will route to ZEISS online shop where you can purchase a formal license.
- Activate: After getting your license, you can enter the license Key here to activate it. Please note, internet connection is required.

17 Application and System Requirements

Application

| Category / Field of Appli- cation | Education, Documentation, Microphotography, Laboratory and Research |
|--------------------------------------|---|
| Compatible ZEISS micro- scopes | Primostar 3 HD, Primo Star HDcam, Primotech (Windows and iOS only), Primovert HDcam, Stemi 305 cam, Axioscope 5/7/ Vario and Axiolab 5 |
| Compatible ZEISS cameras | Axiocam ERc 5s, Axiocam 202 mono, Axiocam 208 color, Ax- iocam 105 color (Windows only) and Axiocam 305 mono/ color (Windows only) |
| Languages | English, Czech, French, German, Italian, Japanese, Korean, Polish, Portuguese, Russian, Simplified Chinese, Spanish |

System Requirements

| | Labscope v4.0 for Android | Labscope v3.3 for iPhone | Labscope v3.3 for iPad | Labscope v3.4 for Windows |
|--|------------------------------|---------------------------------------|--|--|
| Operating Sys- tem | Android 9.0 or later | iOS 13 or later | iOS 13 or later | Windows 10 (64-bit) |
| Minimum Hardware Re- quirements | RAM: 4GB | iPhone 6s/plus iPhone SE (2016) | iPad Air 2 iPad 5 iPad Mini 4 | CPU: i3 (6th generation) dual-core @ 2.5GHz RAM: 4GB |
| Recom- mended Hard- ware Require- ments | RAM: 6GB or above | iPhone 7 or later | iPad Air 3 or later iPad 6 or later iPad Mini 5 or later iPad Pro 1 or later | CPU: i5 (7th generation) quad-core @ 3.0GHz or above RAM: 8GB or above |

System requirements ZEISS Labscope Module Fast Panoram

| Operating system | Windows 10 (64-bit) |
|----------------------|--|
| Minimum hardware | CPU: i5 (8th generation) quad-core @ 3.0 GHz RAM: 8 GB or above |
| Recommended hardware | CPU: i7 (9th generation) hexa-core @ 4.0 GHz or above RAM: 16 GB or above |

Note:

Supported camera types for Multi Channel module: Axiocam 105 color, Axiocam 202 mono, Axiocam 208 color and Axiocam 305 mono/color. Supported camera types for Fast Panorama module: Axiocam 305 mono/color.

Not all performance specifications of the used cameras can be supplied in the Labscope environment due to application specific restrictions.

Carl Zeiss Microscopy GmbH

Carl-Zeiss-Promenade 10 07745 Jena Germany phone: +49 3641 64 3161 fax: +49 3641 64 3439 info.microscopy@zeiss.com www.zeiss.com/microscopy